Functionality Transfer from C₈ to C₉ in Sesquiterpenes. Synthesis of the named Herbolide E from Artemisin.

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Abstract: The key intermediate 6 was obtained in five steps from artemisin (1). The spectroscopic characteristics of the synthetic product 7a reveal that the proposed structure for the natural herbolide E must be revised.

Sesquiterpene lactones constitute a group of natural compounds widely distributed in the vegetal kingdom, which exhibit a broad spectrum of biological activities. In the last years a number of 9-oxyfunctionalized sesquiterpene lactones have been isolated from natural sources.¹ However efficient methodology to functionalize C-9 has not yet been reported.

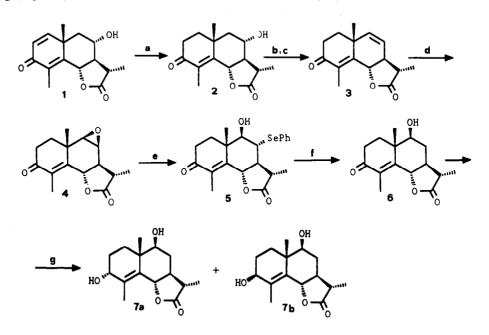
In continuation of our research programme on synthesis of natural sesquiterpene lactones² we report in this *Letter* a functionality transfer from C8 to C9 in an eudesmane framework obtaining the 9-hydroxy-6,12-eudesmanolide 6 starting from artemisin (1). As a first example of the synthetic utility of this functionality transfer we also report in this *Letter*, the transformation of 6 into the named herbolide E (7a).

In our approach to the desired 9-oxyfunctionalization we regard the 8,9-epoxide 4 as the key product. So, the starting material 1 was converted, by catalytic hydrogenation over Wilkinson catalyst,³ into 1,2-dihydroartemisin (2), which was dehydrated to alkene 3. Several attempts of dehydration of hydroxyl at C₈ through the chloride, mesylate and sulfoxide were unsuccessful. However, conversion of 2 into its triflate (triflic anhydride, pyridine/CH₂Cl₂)⁴ followed by elimination with Li₂CO₃/ N,N-dimethylacetamide⁵ afforded 3 (yield 52%).⁶

Inicial experiments of epoxidation of this alkene with *m*-chloroperbenzoic acid,⁷ hexafluoroacetone-H₂O₂⁸ or sodium perborate-acetic anhydride⁹ were unsuccessful as they gave rise to the recovery of unreacted starting product only. However, treatment of this alkene 3 with dimethyldioxirane¹⁰ provides the epoxide 4^{11} in excellent chemo- and stereoselectivity and yield (99%).

In order to open the oxirane ring, compound 4 was treated with different reagents [PhSH/HNa, PhSH/Ti(*i*-PrO)4].¹² Successful result was obtained with PhSeNa/Ti(*i*-PrO)4/DMF¹³ yielding 5 (85%).¹⁴ Once the oxygen function was transferred from C8 to C9, the elimination of the phenylselenyl group was carried out by treatment with Raney Ni¹⁵ giving the compound 6 (80%).¹⁶

This product 6, with a hydroxyl group at C9, constitute a key intermediate in the synthesis of 9-oxyfunctionalizated sesquiterpenoids with diverse functionalization at the A ring.¹⁷ In this communication we report its NaBH4 reduction, which afforded the epimeric alcohols $7a^{18}$ and $7b^{19}$ (1:3 ratio) in good yield (84%).



Reagents and conditions: a) H₂, benzene, Wilkinson Cat., overnight; b) Triflic anhydride (1.5 mmol), pyridine (1.5 mmol), CH₂Cl₂, -15° C, 30 min. c) Li₂CO₃ (8 mmol), N,N-dimethylacetamide, 60° C, 30 min. d) Dimethyl dioxirane (1.2 mmol), acetone, CH₂Cl₂, 0° C, 9 h. e) Ph₂Se₂ (2.5 mmol)-NaBH₄ (2.7 mmol), AcOH (1 mmol), Ti(*i*-PrO)₄ (1 mmol), DMF, r.t., 25 h. f) Deactivated Raney Ni, MeOH, r.t. 15 min. g) NaBH₄ (16 mmol), MeOH, 0° C, 5 min.

The ¹H spectrum of the 3α -alcohol 7a showed a broad singlet at δ 3.90 for H₃ in equatorial position, a double doublet at δ 3.47, J 11.5 and 4.6 Hz for H₉, indicating the β -equatorial disposition of the hydroxyl group at this carbon and a double doublet at δ 4.51, J 11.0 and 1.2 Hz for the lactonic proton (H₆). These signals, as all the remaining signals of the ¹H NMR spectrum, were assigned by spin decoupling. As in the starting artemisin the C₁₁-methyl group is α . This point was verified from the coupling J_{7,11} observed in the signal of H₁₁ which appeared as a double quartet at δ 2.29 (J 7,11 = 11.5 and J 11,13 = 6.5 Hz), as well as by the chemical shift (δ 12.4) of the C₁₃ in the ¹³C NMR spectrum.

These data are consistent with the structure 7a, which has been proposed for herbolide E^{20} a metabolite isolated from *Artemisia herba alba* growing in Israel, and which structure was elucidated by spectroscopic methods in 1984. However the ¹H and ¹³C NMR spectra described for this natural product do not coincide with those of the synthetic product, especially in the δ values of H₆, H₇ and H₉ which

appear at δ 4.51, 1.70 and 3.47 in the synthetic product and at δ 4.82, 2.27 and 3.82 in the natural product, and in the ¹³C NMR values for C₁₃ at δ 12.4 in the synthetic product and δ 9.9 in the natural product. The downfield shift of H₆ and H₇ signals as well as the δ value for C₁₃ in the natural product are in good agreement with a β -methyl group at C₁₁^{21,22} and consequently we think that the structure of the natural herbolide E might be the C₁₁-epimer of compound **7a**.

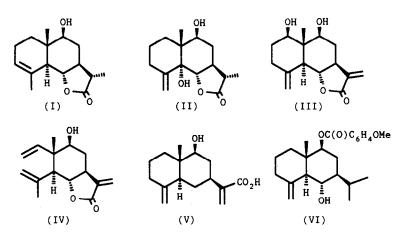
Further synthetic studies on other herbolides are in progress.

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- 11. Compound (4): m.p. 185-197°C, dec. (Hexane-EtOAc); $[\alpha]D + 41.3$ (CHCl3); IR(KBr) 3020, 1780, 1665 cm⁻¹; ¹H NMR (200 MHz, CDCl3) δ 1.38 (3H, d, J 6.7 Hz, H13), 1.44 (3H, s, H14), 1.93 (3H, d, J 1.3 Hz, H15), 2.10 (1H, dt, J 6.0, 13.0 Hz, H1 α), 2.32 (1H, dd, J 11.0, 12.7 Hz, H7), 2.81(1H, qd, J 6.7, 12.7 Hz, H11), 2.93 (1H, d, J 3.8 Hz, H9), 3.54 (1H, d, J 3.8 Hz, H8), 4.86 (1H, dd, J 1.0, 11.0 Hz, H6).
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- Compound (5): m.p. 167-168°C (Hexane-EtOAc); [α]D + 72.9 (CHCl3); IR(KBr) 3560-3300, 1780, 1670, 1620, 920, 735, 690 cm⁻¹; ¹H NMR (200 MHz, CDCl3) δ 1.33 (3H, s, H14), 1.61 (3H, d, J 6.8 Hz, H13), 1.94 (3H, d, J 1.0 Hz, H15), 2.03 (1H, t, J 11.0 Hz, H7), 2.25 (1H, dt, J 4.3, 13.5 Hz, H1B), 2.57 (1H, qd, J 6.8, 11.0 Hz, H11), 3.15 (1H, d, J 11.0 Hz, H9), 3.22 (1H, t, J 11.0 Hz, H8), 4.67 (1H, dd, J 1.0, 11.0 Hz, H6), 7.4-7.2 (3H, m, Ar), 7.58 (2H, dd, J 2.0, 7.5 Hz, Ar).

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- 16. Compound (6): m.p.167-169°C (Hexane-EtOAc); $[\alpha]D + 64.1$ (CHCl₃); IR(KBr) 3560-3200, 1775, 1660, 1620 cm⁻¹; ¹H NMR (200 MHz, CDCl₃) & 1.28 (3H, d, J 7.3 Hz, H₁₃), 1.30 (3H, s, H₁₄), 1.72 (1H, q, J 11.5 Hz, H₈₈), 1.87 (1H, dt, J 3.4, 11.5 Hz, H₇), 2.01 (3H, d, J 1.2 Hz, H₁₅), 2.15 (1H, td, J 4.8, 13.2 Hz, H₁₆), 2.22 (1H, ddd, J 3.4, 4.6, 11.5 Hz, H_{8α}), 2.39 (1H, qd, J 7.2, 11.5 Hz, H₁₁), 3.60 (1H, dd, J 4.6, 11.5 Hz, H₉), 4.67 (1H, dd, J 1.2, 11.0 Hz, H₆).
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- Compound (7a): an oil, [α]D +44.0 (CHCl3); IR(NaCl) 3540-3200, 1780, 1665 cm⁻¹; ¹H NMR (200 MHz, CDCl3) δ 1.08 (3H, s, H14), 1.22 (3H, d, J 6.9 Hz, H13), 1.58 (1H, q, J 11.5 Hz, H8B),1.6-1.9 (5H, m, 2 H1, 2 H2, H7), 2.00 (3H, d, J 1.2 Hz, H15), 2.11 (1H, ddd, J 2.5, 4.6, 11.5 Hz, H8α), 2.29 (1H, qd, J 6.9, 11.5 Hz, H11), 3.47 (1H, dd, J 4.6, 11.5 Hz, H9), 3.90 (1H, brs, H3), 4.51 (1H, dd, J 1.2, 11.0 Hz, H6). ¹³C RMN (50,3 Mz, CDCl3) δ 178.3 (C12), 132.2 (C4 or C5), 129.4 (C5 or C4), 81.7 (C6), 78.6 (C3), 70.3 (C9), 48.8 (C7), 42.3 (C10), 40.8 (C11), 32.4 (C1 or C8), 31.2 (C8 or C1), 27.1 (C2), 17.9 (overlapped signals, C14 and C15), 12.4 (C13).
- 19. Compound (**7b**): m.p.177-179°C (Hexane-CH₂Cl₂); $[\alpha]D$ -31.1 (CHCl₃); IR(KBr) 3540-3120, 1760, 1640 cm⁻¹; ¹H NMR (200 MHz, CDCl₃) δ 1.10 (3H, s, H₁₄), 1.22 (3H, d, J 6.9 Hz, H₁₃), 1.93 (3H, d, J 1.2 Hz, H₁₅), 2.11 (ddd, J 2.4, 4.8, 11.5 Hz, H_{8 α}), 2.27 (1H, qd, J 6.9, 11.7 Hz, H₁₁), 3.45 (1H, dd, J 4.8, 10.7 Hz, H9), 3.90 (1H, brt, J 5.1, H₃), 4.54 (1H, dd, J 1.2, 10.5 Hz, H₆).
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